



# Optimization of Temperature and Extraction Time for the Nutritional And Phytochemical Contents of Corn Bran With Ultrasonic-Method

H Haslina<sup>1</sup>, D Larasati<sup>1</sup>, E Pratiwy<sup>1</sup>, N Nazir,<sup>2</sup> and I Fitriana<sup>1</sup>

<sup>1</sup> Faculty of Agriculture Technology, Semarang University, Semarang 50196, Central Java, Indonesia

<sup>2</sup> Faculty of Agricultural Technology, Andalas University, Padang 26123, Indonesia

Corresponding author: [chana\\_panca@yahoo.com](mailto:chana_panca@yahoo.com)

## ARTICLE INFO

### Article History::

Received: 11 August 21

Final Revision: 26 August 21

Accepted: 21 October 21

Online Publication: 25 October 21

## KEYWORDS

Corn bran, extract, nutritional, phytochemical, ultrasonic-method

## CORRESPONDING AUTHOR

\*E-mail: [chana\\_panca@yahoo.com](mailto:chana_panca@yahoo.com)

## ABSTRACT

One of the natural antioxidants sources that have the potential to be utilized as raw material for functional food is corn bran. Corn bran is a by-product of the milling process of corn that is gaining attention as a functional food is increasing in recent years. This study aimed to optimize temperature and extraction time for the nutritional and phytochemical contents of corn bran with ultrasonic-method. Optimized Custom Design was applied to investigate the effect of experimental factors on the nutritional and phytochemical contents. This study used Randomized Block Design (RBD) arranged in-factorial with 3 treatments, namely temperature and time of extraction. Temperature: A1=50°C, A2=55°C, A3=60°C, and A4=65°C, and time: B1=10 minutes, B2=15 minutes, B3=20 minutes, and B4=25 minutes. The data obtained were analyzed statistically using ANOVA with a significance level of 95% and then processed with Software DX13.0 © Program. The results of the research show the formula of the experiment which is optimal at a temperature of 50°C and 10 minutes. In this condition, the result is the yield at 38.34%, nutritional contents (water at 9.17%; ash at 0.33%; fat at 1.33%, protein at 4.40%, carbohydrates at 85.47%; and crude fiber at 1.88%. produce yield 38.34%, and phytochemical contents (total phenols at 1778.07 µg GAE/g, flavonoids at 92.11 µg GAE/g, vitamin C at 5.84 mg, antioxidant activity at 43.33%, and tannins at 0.11%). This study implies that there is an increase in added value from the conversion of corn bran into nutrient-rich products and has a promising phytochemical content.

## 1. INTRODUCTION

### 1.1. Research Background

Zea comes from ancient Greek and is a generic name for cereal and grains. Corn (*Zea mays* L.) is the world's third-leading cereal crop, after wheat and rice [1]. Corn (*Zea mays* L.) is a potential source of carbohydrates like flour and corn starch. The demand for corn is constantly increasing for food, fuel, and feed [2]. Corn is a plant that is resistant to and adaptable to environmental conditions. The growth and development of corn depend on the ecological characteristics of the external conditions, the longevity of the species, water demand, soil temperature, and air temperature, light, and nutrient supply [3].

Rice bran is a by-product of the milling of rice paddy, which is a brown fine powder from the outermost layer of the cracked rice paddy [1]. The results of the rice milling are to produce rice (70%), rice bran (8%), and embryo (2%) [2]. Corn bran is a mixture of the aleurone layer and pericarp that

regardless in the process of grinding corn. The process of grinding and corn milling will produce 16 to 28% husks (hulls), 6-11% bran, 2-4% polish, and about 60 percent of the endosperm. Corn bran is also rich in nutrients. As the nutritional content consists water at 1%; calories at 356; protein at 9%; fat at 8.5%; carbohydrate at 64.5%; Ca at 200 mg; Fe at 10 mg; P at 500 mg; vitamin A at 51 mg; vitamin B at 1.2 mg, and vitamin C at 89% [3]. During this time, they are used as cattle feed because not many people know the health benefits. When the level of rice bran is about 10% [4], then the potential of rice bran that can be utilized is about 347 tons. This amount is quite large and has the potential to be used as one of the raw materials of the food industry. Bran is reported to contain some phenolic compounds, antioxidants, and is rich in dietary fiber, vitamins, and minerals [5], [6]. In addition, it is a source of dietary fiber, especially do not dissolve, contribute to the increased levels of iron and zinc, as a product of low-calorie and low-phytic, negative effect on the bioavailability of iron in rats, and can be used for the enrichment of food products [7]. Some research on the functionality of bran for health, among others: anticancer, antihypcholesterolemic, and antiatherogenic [5], [8]–[11].

## 1.2. Literature Review

Bioactive components contained in a material can be obtained by the method of extraction. There are various extraction methods, namely maceration, percolation, ultrasonic, and soxhlet. The extraction method used in this research is the method of extraction with ultrasonic waves. The method of extraction with the use of ultrasonic waves that acoustic waves with a frequency greater than 16 kHz. The method of extraction with ultrasonic waves is known to have advantages compared with the method of maceration. One of the advantages of the method of extraction ultrasonic is the speed of extraction, compared with extraction by thermal or conventional. The method of extraction with the ultrasonic waves is safer, shorter, and increases the yield of coarse [12], [13]. Factors that can affect the extraction temperature and the extraction time used. Research reported an increase in the temperature in the extraction process needs to be considered, extraction temperature is too high, and the extraction time is too long and exceeds the optimum limit can lead to loss of compounds in solution because of the occurring process of oxidation [14]. Bioactive components such as flavonoids are not resistant to a high temperature above 50°C, which changes the structure as well as produces an extract that low. Extraction temperature is too low, and extraction time is too short will cause the bioactive components extracted from the material is not the maximum so that the bioactive components obtained low [15]. A study about the influence of temperature and time of extraction of the leaves of the soursop using the ultrasonic that the temperature of 45°C with a time of 20 minutes gives the best result with the value of the yield, total flavonoids, and high antioxidant activity [15].

One of the solvents that are often used for the extraction of flavonoid compounds is ethanol. Ethanol including a polar solvent is safe, easily available, and environmentally friendly. Ethanol also called ethyl alcohol is a chemical substance that belongs to the class of alcohol [16]. Ethanol has a chemical structure  $\text{CH}_3\text{CH}_2\text{OH}$ , has the contents of a volatile, colorless, and are polar so it is used as a solvent for many compounds [17]. According to Ref. [18], several contributing factors affect the rate and quality of the extraction on the components of the bioactive compounds phenol, namely the type of extraction solvent, solvent concentration, particle size, temperature, pH, and extraction time. Meanwhile, according to Ref. [19], the factors that affect the rate of extraction are the surface area, a comparison of solute and solvent, speed, and time of stirring.

## 1.3. Research Objective

This study aimed to optimize temperature and extraction time for the nutritional and phytochemical content of corn bran with ultrasonic-method.

## 2. MATERIALS AND METHODS

### 2.1. Materials

The raw material in this study is corn bran varieties NK212, age 120 days, obtained from corn farmers in Mranggen, Demak Regency, Central Java, Indonesia, and the chemical materials used to test the nutritional and phytochemical content obtained from the Laboratory of Chemistry and Biochemistry the

Department of Agricultural Technology, Semarang University, Indonesia.

### 2.2. Equipment

The tools used for producing corn bran powder include a knife, pan, blender, dry (National PBL-104), cabinet dryer automatic (OVG-12), sieve 60 mesh (ATE-126, 0.250 mm), thermometer, plastic, and brush. The tools used for the extraction of corn bran powder is the ultrasonic vibrating device (Cole Palmer/CPX 130), stirrer glassware (Pyrex), smooth filter paper, rotary evaporator (Buchi B-490), freezer, vacuum filter (Butchi V500), shaker water bath, analytical balance, pipette drops, dark bottle, nitrogen spray equipment, and aluminum foil. The tools used for the analysis include the stove, oven electric, pH meter, analytical balance, color reader, becker glass, petri dish, desiccator, suction ball, pipette, lamp, thermometers, racks, aluminum foil, wooden tube, cuvette, spectrophotometer UV-Vis, Folin-Ciocalteu colorimetric, centrifuge, GC-MS, and some glassware for analysis.

### 2.3. Preparation of Materials

Fresh corn bran washed with distilled water, dried by oven at a temperature of 60°C for 24 hours [20] the final water content is 10 %, pounded into powder using a grinder, then vacuum packaged and stored <-20°C until it is used for analysis.

### 2.4. Extraction of Corn Bran

The powder of corn bran weighed as much as 15 grams with an analytical balance, put into an Erlenmeyer flask. Added ethanol 70% as much as 150 ml (1:10), then extracted with a combination of a temperature of 50°C, 55°C, 60°C, and 65°C with a time of 10, 15, 20, and 25 min using an ultrasonic bath. Powder bran corn that has been extracted with ultrasonic bath and then filtered using Whatman paper no.1. The filtrate obtained was then performed evaporation. Evaporation is done by a rotary vacuum evaporator with a pressure of 100 bar, a temperature of 40°C, and 100 rounds rpm [21] that have been modified). The condensed extract obtained was weighed and calculated the yield of the extract was then placed in a bottle, for further analysis.

### 2.5. Analysis

Analysis of the nutritional and phytochemical contents carried out on water content and ash using the oven method [22], the protein content using the method of Micro-Kjeldahl [22], the fat content using Soxhlet method [22], and the carbohydrate content using carbohydrate by difference method [22], and the crude fibers [22], as well as the phytochemicals contents (total phenol [23], total flavonoids [24], vitamin C [25], antioxidant activity [23], tannins [24], and yield.

### 2.6. Experimental Design

The experimental design used in this study is a Randomized Block Design (RBD) factorial with 3 replications. The treatment used is the temperature of extraction (A1=50°C, A2=55°C, A3=60°C, and A4=65°C). The time of extraction (B1=10 minutes, B2=15 minutes, B3=20 minutes, and B4=25 minutes). Furthermore, the data obtained were analyzed statistically using ANOVA with a significance level of 95 % then processed with the Software Design Expert 13 ® program with Response

Surface Methodology (RSM) to determine the optimum concentration of the extract powder of corn bran. After a

combination of 2 treatments which is the temperature and the time of extraction were shown in Table 1.

Table 1. Independent variables

Component	Independent variable	Category	Minimal	Maximal
X1	Temperature	50°C, 55°C, 60°C, and 65°C	50	65
X2	Time	10, 15, 20, and 25 minutes	10	25

The data are expressed as mean and standard deviation (SD). Furthermore, the data obtained were analyzed statistically using ANOVA with a significance level of 95% and if there is a difference between the treatment was continued with Duncan New Multiple Range Test (DNMRT) at the level of 5%. The results of the research were processed with the software

Design's DX 13 ® to determine the optimum concentration of temperature and time extraction from corn bran.

### 3. RESULT AND DISCUSSION

#### 3.1. Description of nutritional content

Table 2. Description of nutritional content

Treatment	Components						
	Yield (%)	Water (%)	Ash (%)	Fat (%)	Protein (%)	Carbohydrates (%)	Crude fibers (%)
A1B1	38.40 ±0.46	9.77 ±0.10	0.33 ±0.00	1.38 ±0.02	4.41 ±0.05	84.16 ±0.17	1.88 ±0.02
A1B2	37.88 ±0.26	7.47 ±0.52	0.32 ±0.00	1.40 ±0.01	4.35 ±0.03	86.46 ±0.47	1.84 ±0.01
A1B3	37.73 ±0.45	6.97 ±0.13	0.32 ±0.00	1.44 ±0.02	4.33 ±0.05	86.95 ±0.20	1.85 ±0.02
A1B4	37.07 ±0.06	6.23 ±0.16	0.30 ±0.00	1.46 ±0.00	4.26 ±0.01	87.75 ±0.17	1.85 ±0.00
A2B1	37.07 ±0.44	7.34 ±0.38	0.32 ±0.00	1.25 ±0.01	4.18 ±0.05	86.92 ±0.31	1.82 ±0.02
A2B2	36.29 ±0.25	5.91 ±0.18	0.31 ±0.00	1.35 ±0.01	4.09 ±0.03	88.35 ±0.15	1.75 ±0.01
A2B3	35.90 ±0.43	5.68 ±0.31	0.30 ±0.00	1.42 ±0.02	4.04 ±0.05	88.57 ±0.24	1.74 ±0.02
A2B4	34.76 ±0.05	5.26 ±0.17	0.28 ±0.00	1.44 ±0.00	3.92 ±0.01	89.11 ±0.18	1.70 ±0.00
A3B1	35.14 ±0.42	7.22 ±0.15	0.31 ±0.00	1.25 ±0.01	4.04 ±0.05	87.18 ±0.21	1.80 ±0.02
A3B2	34.64 ±0.24	5.53 ±0.08	0.31 ±0.00	1.32 ±0.01	3.98 ±0.03	88.87 ±0.04	1.74 ±0.01
A3B3	33.98 ±0.41	5.35 ±0.22	0.30 ±0.00	1.37 ±0.02	3.90 ±0.05	89.07 ±0.16	1.76 ±0.02
A3B4	32.81 ±0.05	4.54 ±0.06	0.28 ±0.00	1.41 ±0.00	3.77 ±0.01	90.01 ±0.07	1.72 ±0.00
A4B1	30.83 ±0.37	5.51 ±0.11	0.31 ±0.00	1.20 ±0.01	3.44 ±0.04	89.55 ±0.16	1.78 ±0.02
A4B2	30.32 ±0.21	5.10 ±0.23	0.30 ±0.00	1.25 ±0.01	3.40 ±0.02	89.96 ±0.20	1.75 ±0.01
A4B3	29.75 ±0.35	4.45 ±0.48	0.29 ±0.00	1.28 ±0.02	3.33 ±0.04	90.65 ±0.43	1.72 ±0.02
A4B4	28.17 ±0.04	4.51 ±0.12	0.26 ±0.00	1.29 ±0.00	3.14 ±0.01	90.80 ±0.12	1.63 ±0.00

Description:

Temperatur: A1 (50°C), A2 (55°C), A3 (60°C), A4 (65°C)

Time: B1 (10 minutes), B2 (15 minutes), B3 (20 minutes), B4 (25 minutes)

#### 3.1.1 Yield

Fig. 1 the contour plot of the yield of a dark blue color indicates the response of the lowest yield at 28.17% and the dark red color shows the response of the highest yield at 38.40%. The lines consisting of dots on the chart contour plot show the combination of these two components with a different number that produces the same response of yield. Table 2 shows that the yield obtained in A4B4 (65°C and the temperature-time 25 minutes) at 28.17%, while the highest average soluble fiber is achieved on A1B1 (50°C and the temperature is 10 minutes) at 38.40%.

The lowest average yield was obtained in A4B4 (65°C and the temperature 25 minutes) at 28.17%, while the highest average soluble fiber was achieved on A1B1 (50°C and the temperature is 10 minutes) at 38.40%. The different treatment causes different yield. The difference in the suspected presence of evaporation leads to reduced ability of the yield of corn bran. From Table 2 it is known that the higher the temperature and the longer the extraction time, the yield of the extract produced will be lower. This is by following per under research conducted by [26], [27]. The higher the temperature and the longer the time of extraction, the lower the yield obtained. This is due to too high

temperatures with a long time cause more water content evaporated, resulting in yield decrease, and vice versa, the lower the temperature used, then the less water evaporated so it can obtain the high yield [28]–[31].

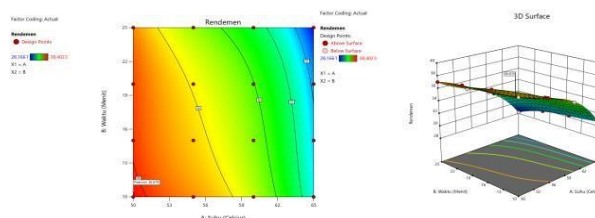


Fig. 1. Contour plot chart and 3D surface yield

#### 3.1.2 Water content

Fig. 2 chart contour plot of the value of the water content of the dark blue color indicates the value of the response of the lowest water content at 4.45% and a dark red color shows the response of the highest water content at 9.77%. The lines consisting of dots on the chart contour plot show the combination of these two components with a different number that produces the same response of the water content. Table 2

shows that, the lowest average of water content obtained on the parameters of A4B4 (65°C and the temperature-time 25 minutes) at 4.51%, while the highest average of water content is obtained in the A1B1 (50°C and the temperature is 10 minutes) at 9.77%. Fig. 2 shows that with the high temperature and long extraction time, then the water content in the corn bran will decrease.

The lowest average of water content is obtained on the parameters of A4B4 (65°C and the temperature-time 25 minutes) at 4.51%, while the highest average of water content is obtained in the A1B1 (50°C and the temperature is 10 minutes) at 9.77%. Figure 2 shows that the high temperature and long extraction time, then the water content in the corn bran will decrease. These results are by following per with the results of previous research conducted by [32]. According to [33], the longer the cooking time, the more water content will decrease, causing the evaporation of more water, so the water content in the material is vanishingly small. This is due to the higher extraction temperature and the longer the presence of heat in the extraction process, the greater the heat energy that will carry the air, so that the more the amount of mass of water evaporated from the surface of the material [34]. Evaporation is also caused due to the occurrence of the difference in vapor pressure between the water on the material with water vapor in the air. The water vapor pressure on the material, in general, is greater than the vapor pressure of water in the air, resulting in the displacement of the water mass of material into the air [35].

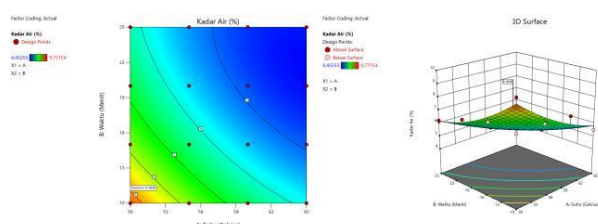


Fig. 2. The chart contour plot and 3D surface water content

### 3.1.3 Ash content

Fig. 3 chart contour plot of the value of the ash content of the dark blue color indicates the value of the response of the lowest ash content at 0.26% and a dark red color shows the response of the highest ash content at 0.33%. The lines consisting of dots on the chart contour plot show the combination of these two components with a different number that produces the same response of ash content. The lowest average of ash content was obtained in A4B4 (65°C and the temperature-time 25 minutes) at 0.26%, while the highest average ash content is obtained in the A1B1 (50°C and the temperature is 10 minutes) at 0.33%. From table 2 it is known that the higher the temperature and the longer the extraction time, the more ash content will decrease.

The lowest average of ash content was obtained in A4B4 (65°C and the temperature-time 25 minutes) at 0.26%, while the highest average ash content was obtained in the A1B1 (50°C and the temperature is 10 minutes) at 0.33%. From table 2 it is known that the higher the temperature and the longer the extraction time, the more ash content will decrease. According to [36], this is presumably due to the longer and higher the temperature of extraction used will make the lower ash content, due to the higher the water that out of the material surface. The value of ash content in the corn bran can be derived from the

content of mineral material [37], [38]. The greater the ash content of a food material showed higher mineral that was conceived by the food [39].

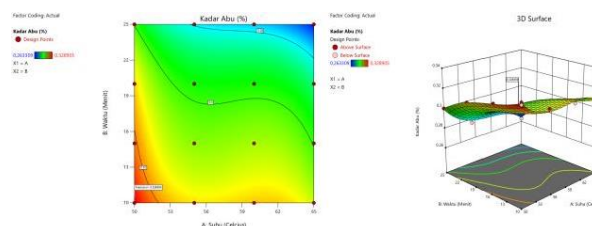


Fig. 3. Contour plot chart and 3D surface ash content

### 3.1.4 Fat content

Fig. 4 chart of the contour plot of the value of the fat content of dark blue color indicates the value of the response of the lowest average of fat content at 1.20% and a dark red color shows the response of the highest average of fat content at 1.46%. The lines consisting of dots on the chart contour plot shows the combination of these two components with a different number that produces the same response of the fat content. The lowest average of fat content was obtained in A4B1 at 1.20%, while the highest average of fat content was obtained on A1B4 at 1.46%. From Table 2 it can be known that the fat content increased. The lowest average protein content obtained in A4B4 (65°C and the temperature-time 25 minutes) at 3.14%, while the highest average of protein content is obtained in the A1B1 (50°C and the temperature is 10 minutes) at 4.41%.

The lowest average of fat content was obtained in A4B1 at 1.20%, while the highest average of fat content was obtained on A1B4 at 1.46%. From Table 2 it can be known that the fat content increased. The fat content will increase directly proportional with the increase of the time and temperature of extraction. This means that the longer the time of extraction with the higher temperature, will cause the contact between the solute and solvent gets long so that the fat content obtained will be higher [40]. Decreased levels of fat in the material allegedly because it contains free fatty acids, in which the free fatty acid has a solubility pretty good in ethanol [41]. Low-fat content can be caused by the high water content so that the fat content is proportionally decreased [42].

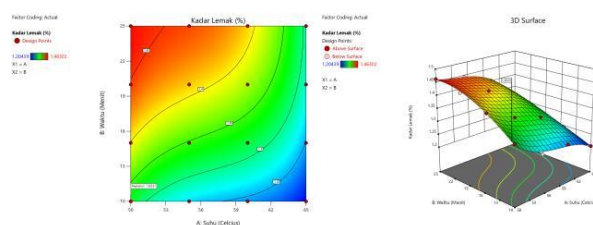


Fig. 4. Contour plot chart and 3D surface fat content

### 3.1.5 Protein content

Fig. 5 chart contour plot of the value of protein content of the dark blue color indicates the value of the response of the lowest average of protein content at 3.14% and a dark red color shows the response of the highest average of protein content at 4.41%. The lines consisting of dots on the chart contour plot show the combination of these two components with the



different amounts that produce the same response of the protein content.

The lowest average protein content obtained in A4B4 (65°C and the temperature-time 25 minutes) at 3.14%, while the highest average of protein content is obtained in the A1B1 (50°C and the temperature is 10 minutes) at 4.41%. With the increase of temperature and time extraction, the protein content in corn bran will decrease. This is because the higher the temperature used causes the protein in the extract to decrease. After all, because protein is a nutrient that is easily damaged when it is exposed to high heat [43], [44]. According to [45], the solubility of the protein generally increased when the temperature rises 0-40°C, and the longer the time of the dissolution, then the more contact between the solute and solvent is getting long, so the solute that is taken will be higher.

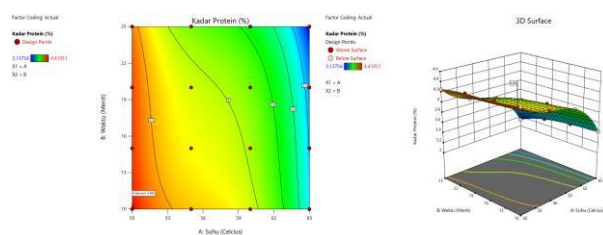


Fig. 5. Contour plot chart and 3D surface protein content

### 3.1.6 Carbohydrate content

Fig. 6 chart of the contour plot of the value of the carbohydrates content, dark blue color indicates the value of the response of the lowest average of carbohydrate content at 84.16%, and a dark red color shows the response of the highest average of carbohydrate content at 90.80%. The lines consisting of dots on the chart contour plot show the combination of these two components with a different number that produces the same response of the carbohydrates content. The lowest average of carbohydrate content was obtained in the A1B1 (50°C and the temperature is 10 minutes) at 84.16%, while the highest average of carbohydrate content was obtained on A4B4 (65°C and the temperature-time 25 minutes) at 90.80%.

The lowest average of carbohydrate content was obtained in the A1B1 (50°C and the temperature is 10 minutes) at 84.16%, while the highest average of carbohydrate content was obtained on A4B4 (65°C and the temperature-time 25 minutes) at 90.80%. The longer the extraction time, the carbohydrate content will be higher, this is presumably due to the higher the total sugar content in the extract [46]. It is also supported by a statement from [47], the presence of water evaporation during heating, causing the water content to decrease and the concentration of solids will increase. The decline in water content will also make the higher levels of nutrients that are left behind, including carbohydrates [48].

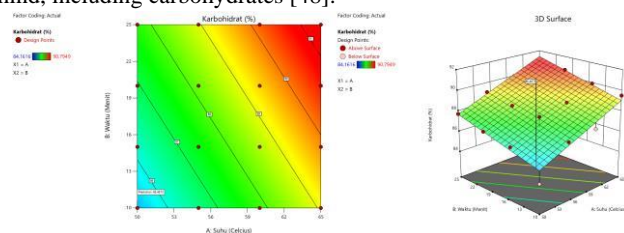


Fig. 6. Contour plot chart and 3D surface carbohydrate content

### 3.1.7 Crude fibers

Fig. 7 chart contour plot of the value of crude fiber, dark blue color indicates the value of the response of the lowest average of crude fiber at 1.63% and a dark red color shows the response of the highest average of crude fiber at 1.88%. The lines consisting of dots on the chart contour plot show the combination of these two components with a different number that produces the same response of the crude fiber. The lowest average of crude fiber obtained in A4B4 (65°C and the temperature-time 25 minutes) at 1.63%, while the highest average of crude fiber is obtained in the A1B1 (50°C and the temperature is 10 minutes) at 1.88%. From Table 1. it can be seen that the higher the temperature of extraction and the longer the extraction time, the result in the crude fiber content of the extract will be lower.

The lowest average of crude fiber obtained in A4B4 (65°C and the temperature-time 25 minutes) at 1.63%, while the highest average of crude fiber is obtained in the A1B1 (50°C and the temperature is 10 minutes) at 1.88%. From Table 1. it can be seen that the higher the temperature of extraction and the longer the extraction time, the result in the crude fiber content of the extract will be lower. It is alleged due to a temperature of 40°C and 50°C enzyme amylase contained in the materials is still actively hydrolyzing carbohydrates, as a result, the content of crude fiber will decrease [49], [50]. A decrease in the content of crude fiber in this study, allegedly due to the breakdown of hemicellulose, resulted in the content of crude fiber because hemicellulose is part of the crude fiber.

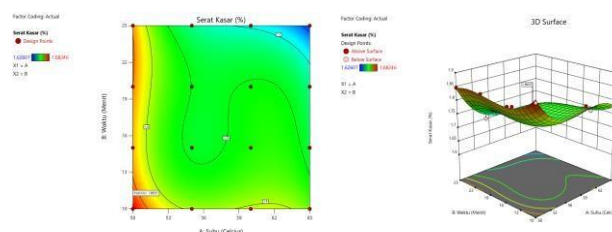


Fig. 7. Contour plot chart and 3D Surface Crude Fibers

## 3.2. Description of phytochemical content

### 3.2.1 Total phenol

Fig. 8 chart contour plot of the value of total phenol, dark blue color indicates the value of the response of the lowest average of total phenols at 1043.18 µg GAE/g and a dark red color shows the response of the highest average of total phenol at 1796.94 µg GAE/g. The lines consisting of dots on the chart contour plot show the combination of these two components with a different number that produces the same response of total phenol. Table 3 indicated that the lowest average of total phenol obtained in the A1B1 of (50°C and the obtained in A4B4 at 1043.18 µg GAE/g (65°C and the temperature-time 25 minutes), while the highest average of temperature is 10 minutes) at 1796.94 µg GAE/g.

The lowest average of total phenol was obtained in the A1B1 of (50°C and the obtained in A4B4 at 1043.18 µg GAE/g (65°C and the temperature-time 25 minutes), while the highest average of temperature is 10 minutes) at 1796.94 µg GAE/g. From Table 3 it is known that the higher the temperature and the longer the extraction time resulted in the decline of total phenols. This is because after passing a temperature of 45°C which is the optimal temperature of total phenol for extraction,

the content of total phenols will decrease [51]. However, at a temperature below 45°C, when the temperature and extraction time is increasing, it will be able to release phenolic compounds of the cell wall or phenolic compounds that are bound, thus causing more and more compounds to be phenol extracted. The use of high temperatures to perform the extraction increases the

solubility of phenol. High temperature can release phenol cell wall or phenolic compound that is bound to that caused by the breakdown of the elements of the cell, thus causing more and more compounds are phenol extracted [52].

Table 3. Description of phytochemical content

Treatment	Components				
	Total phenol ( $\mu\text{g GAE/g}$ )	Flavonoid ( $\mu\text{g GAE/g}$ )	Vitamin C (mg)	Antioxidant activity (%)	Tannins (%)
A1B1	1796.94 $\pm$ 21.42	92.42 $\pm$ 1.10	5.82 $\pm$ 0.07	41.96 $\pm$ 0.50	0.11 $\pm$ 0.00
A1B2	1686.12 $\pm$ 11.51	91.02 $\pm$ 0.62	5.77 $\pm$ 0.04	40.81 $\pm$ 0.28	0.11 $\pm$ 0.00
A1B3	1642.38 $\pm$ 19.58	90.71 $\pm$ 0.62	5.69 $\pm$ 0.07	39.20 $\pm$ 0.47	0.11 $\pm$ 0.00
A1B4	1617.37 $\pm$ 2.40	87.98 $\pm$ 0.13	5.58 $\pm$ 0.01	37.20 $\pm$ 0.06	0.10 $\pm$ 0.00
A2B1	1576.06 $\pm$ 18.79	88.25 $\pm$ 1.05	5.77 $\pm$ 0.07	41.90 $\pm$ 0.50	0.10 $\pm$ 0.00
A2B2	1486.95 $\pm$ 10.15	87.26 $\pm$ 0.60	5.72 $\pm$ 0.04	36.15 $\pm$ 0.25	0.10 $\pm$ 0.00
A2B3	1444.98 $\pm$ 17.22	86.85 $\pm$ 1.04	5.52 $\pm$ 0.07	34.69 $\pm$ 0.41	0.10 $\pm$ 0.00
A2B4	1386.07 $\pm$ 2.06	85.54 $\pm$ 0.13	5.29 $\pm$ 0.01	33.27 $\pm$ 0.05	0.10 $\pm$ 0.00
A3B1	1501.35 $\pm$ 17.90	85.45 $\pm$ 1.02	5.57 $\pm$ 0.07	40.43 $\pm$ 0.48	0.10 $\pm$ 0.00
A3B2	1378.40 $\pm$ 9.41	84.63 $\pm$ 0.58	5.50 $\pm$ 0.04	34.89 $\pm$ 0.24	0.10 $\pm$ 0.00
A3B3	1236.14 $\pm$ 14.73	83.40 $\pm$ 0.99	5.47 $\pm$ 0.07	34.22 $\pm$ 0.41	0.10 $\pm$ 0.00
A3B4	1217.32 $\pm$ 1.81	82.23 $\pm$ 0.12	5.19 $\pm$ 0.01	31.37 $\pm$ 0.05	0.10 $\pm$ 0.00
A4B1	1288.27 $\pm$ 15.36	83.45 $\pm$ 0.99	5.34 $\pm$ 0.06	37.60 $\pm$ 0.45	0.10 $\pm$ 0.00
A4B2	1178.66 $\pm$ 8.05	80.82 $\pm$ 0.55	5.30 $\pm$ 0.04	30.27 $\pm$ 0.21	0.10 $\pm$ 0.00
A4B3	1087.72 $\pm$ 12.97	80.06 $\pm$ 0.95	5.14 $\pm$ 0.06	29.75 $\pm$ 0.35	0.10 $\pm$ 0.00
A4B4	1043.18 $\pm$ 1.55	79.02 $\pm$ 0.12	5.01 $\pm$ 0.01	27.18 $\pm$ 0.04	0.10 $\pm$ 0.00

Description :

Temperatur: A1 (50°C), A2 (55°C), A3 (60°C), A4 (65°C)

Time: B1 (10 minutes), B2 (15 minutes), B3 (20 minutes), B4 (25 minutes)

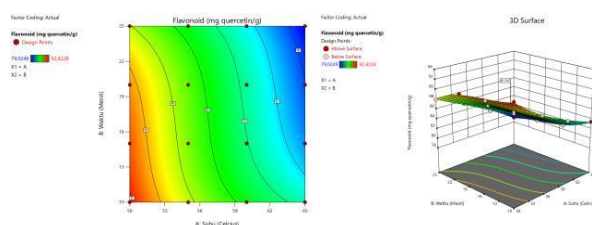


Fig. 8. Contour plot chart and 3D surface total phenol

### 3.2.2 Flavonoids

Fig. 9 chart contour plot of the value of flavonoids dark blue color indicates the value of the lowest average of flavonoids at 79.02  $\mu\text{g GAE/g}$  and a dark red color shows the response of the highest average of flavonoids, which is at 92.42  $\mu\text{g GAE/g}$ . The lines consisting of dots on the chart the contour plot shows the combination of these two components with a different number that produces the same response of the flavonoids. The lowest average of flavonoids was obtained in A4B4 (65°C and the temperature-time 25 minutes) at 79.02  $\mu\text{g GAE/g}$ , while the highest average of flavonoids was obtained in the A1B1 (50°C and the temperature is 10 minutes) at 92.42  $\mu\text{g GAE/g}$ .

The lowest average of flavonoids was obtained in A4B4 (65°C and the temperature-time 25 minutes) at 79.02  $\mu\text{g GAE/g}$ , while the highest average of flavonoids was obtained in the A1B1 (50°C and the temperature is 10 minutes) at 92.42  $\mu\text{g GAE/g}$ . The value of flavonoids in the extract powder of corn

bran is caused by the temperature and time of extraction used. From Table 3 it can be seen that the higher the temperature of extraction and the longer the extraction will make a decline in the content of flavonoids in the extract. The higher the temperature of extraction until it reaches a temperature of 45°C will produce the higher total flavonoids. However, extraction temperature beyond the optimum temperature which is 45°C will cause the total flavonoids to decrease, this is due to the occurrence of the process of oxidation of flavonoid compounds [53]. According to [54], flavonoid compounds are not heat resistant and it is easily oxidized at the high temperature. Flavonoids will be degraded at temperatures above 100°C. Flavonoids are sensitive to heat, because of the hydroxyl groups and ketone, as well as the double bonds of unsaturated [55].

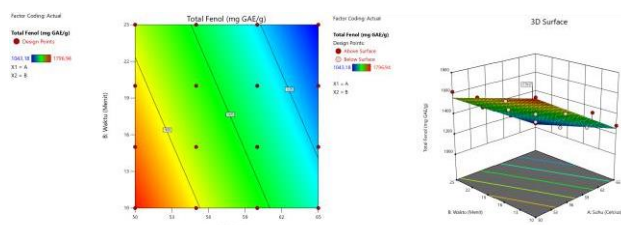


Fig. 9. Contour plot chart and 3D surface flavonoids

### 3.2.3 Vitamin C

Fig. 10 chart contour plot of the value of vitamin C dark blue color indicates the response of the lowest average of vitamin C at 5.01 mg and dark red color shows the response of

the highest average vitamin C, which is at 5.82 mg. The lines consisting of dots on the chart contour plot show the combination of these two components with a different number that produces the same response of vitamin C. The lowest average of vitamin C obtained in A4B4 (65°C and the temperature-time 25 minutes) at 5.01 mg, while the highest average of vitamin C is obtained on A1B1 (50°C and the temperature is 10 minutes) at 5.82 mg. From Table 3 it can be seen that the higher the temperature of extraction and the longer the extraction time will make the vitamin C in the extract is decreasing.

The lowest average of vitamin C was obtained in A4B4 (65°C and the temperature-time 25 minutes) at 5.01 mg, while the highest average of vitamin C is obtained on A1B1 (50°C and the temperature is 10 minutes) at 5.82 mg. From Table 3 it can be seen that the higher the temperature of extraction and the longer the extraction time will make the vitamin C in the extract is decreasing. According to [56], this is because vitamin C is a vitamin that is easily oxidized by heat. Vitamine C is easily damaged when in contact with air (oxidation) especially when it gets heat [57], [58].

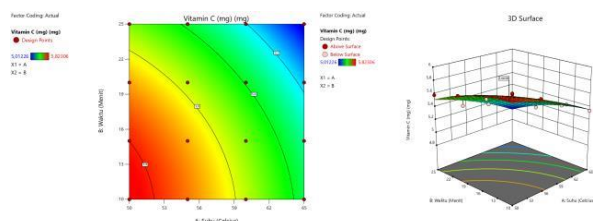


Fig. 10. Contour plot chart and 3D surface vitamin C

### 3.2.4 Antioxidant activity

Fig. 11 chart of the contour plot of the value of the total antioxidant activity, the dark blue color indicates the response of the lowest average of the total antioxidant activity at 27.18%, and a dark red color shows the response of the highest average of total antioxidant activity 41.96%. The lines consisting of dots on the chart contour plot show the combination of these two components with a different number that produces the same response of the antioxidant activity. The lowest average of antioxidants was obtained in A4B4 (60°C and the temperature-time 25 minutes) at 27.18%, while the average of the highest average of antioxidant activity was obtained on the A1B1 (50°C and the temperature is 10 minutes) at 41.96%. It is known that the higher the temperature and the longer the extraction time, the antioxidant activity will decrease (Table 3).

The lowest average of antioxidants was obtained in A4B4 (65°C and the temperature-time 25 minutes) at 27.18%, while the average of the highest average of antioxidant activity was obtained on the A1B1 (50°C and the temperature is 10 minutes) of 41.96%. From Table 3 it is known that the higher the temperature and the longer the extraction time, the antioxidant activity will decrease. The antioxidant activity decreased along with the compounds that are categorized as antioxidants, such as phenolic compounds, flavonoids, and tannins. Phenolic and flavonoids have high antioxidant activity. But after reaching the optimum conditions, then the antioxidant activity will be decreased in the harmony with the decline of compounds that are antioxidants [59]. According to [60] stated that the heating process can extract more antioxidant compounds, but the process of excessive heating will damage the antioxidant

activity. The value of total phenols and flavonoids in the corn bran showed high antioxidant activity. The values of phenol are influenced by its ability to ward off the free radicals [61]. The configuration and the total hydroxyl group are the very basis that affects the mechanism of its activity as an antioxidant. There is a positive correlation between the antioxidant activity with the content of polyphenolic compounds [62].

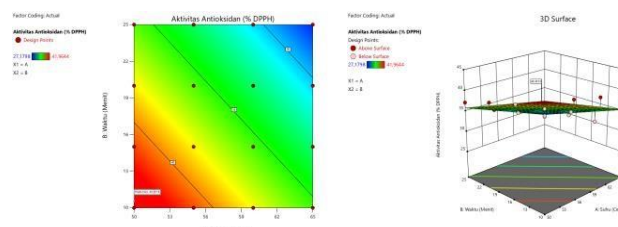


Fig. 11. Contour plot chart and 3D surface antioxidant activity

### 3.2.5 Tannins

Fig. 12 of chart contour plot of the value of the tannins dark blue color indicates the response of the lowest average of tannins at 0.09% and a dark red color shows the response of the highest average of tannins at 0.11%. The lines consisting of dots on the chart contour plot show the combination of these two components with a different number than the same response of tannins. The lowest average of tannins is obtained in A4B4 (65°C and the temperature-time 25 minutes) at 0.09%, while the highest average of tannins is obtained in the A1B1 (50°C and the temperature is 10 minutes) at 0.11%. From Table 3 it can be seen that the content of tannins decreased.

The lowest average of tannins is obtained in A4B4 (65°C and the temperature-time 25 minutes) by 0.09%, while the highest average of tannins is obtained in the A1B1 (50°C and the temperature is 10 minutes) of 0.11%. From Table 3 it can be seen that the content of tannins decreased. The temperature has already reached the optimal point, i.e. the temperature of more than 35°C, tannins will decline. This is because the tannins are damaged from the hydrolysis process during the extraction process and the heating that takes place continuously. Tannins are not resistant to high heat. On the temperature and time that has reached the optimum point then the tannins will decrease, this is because the diffusion process is already not in progress [63].

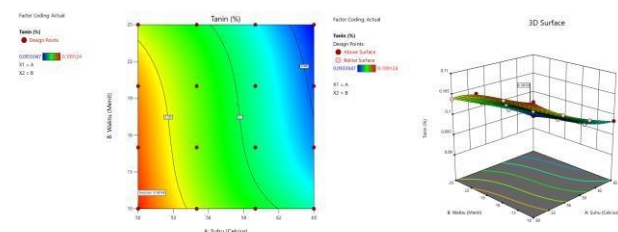


Fig. 12. Contour plot chart and 3D surface tannins

Table 4 shows that the results of the Duncan Test on the difference in the treatment produce the significance on the ANOVA of each of the response levels of water content, ash content, fat content, protein content, carbohydrate, crude fiber, total phenols, flavonoids, vitamin C, antioxidant activity, and tannins in each response no significant difference for each of the factors with a p-value of <0.05 thus there are differences in the

results of the phytochemical testing as a result of the different treatment, which is temperature and time of extraction.

Table 4 shows that the regression coefficient on each response, there is no significant difference for each of the factors with a p-value of  $<0.05$ . The p-value was used to determine the significant presence or absence of each factor. The smaller the p-value, the more significant the price of the coefficient, and the more it contributes to the obtained result. Meanwhile, based on the results of the equations shown, there are no significant differences for all factors with a Predicted R<sup>2</sup> value is more than 0.05. The value of the coefficient price of determination (R<sup>2</sup>) in the model equation was obtained more than 0.05

Table 4. Model analysis for nutritional and phytochemical testing in corn bran

Response	Model	Equation	Significance ( $p < 0.05$ )	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>
Nutritional Content Yield	Cubic	$Y = 35.41 - 2.49A - 0.7149B - 0.3060 AB - 1.47A^2 - 0.3166B^2 + 0.1876A^2B - 0.2636AB^2 - 1.36A^3 - 0.4687B^3$	$< 0.0001$	0.9992	0.9972
Water Content	Quadratic	$Y = 5.50 - 1.28A - 1.10B + 0.4678AB + 0.4463A^2 + 0.5493B^2$	$< 0.0333$	0.9180	0.8419
Ash Content	Cubic	$Y = 0.3016 + 0.0029A - 0.0128B - 0.0035 AB + 0.0049A^2 - 0.0057B^2 - 0.0003A^2B - 0.0022AB^2 - 0.0148A^3 - 0.0058B^3$	$< 0.0243$	0.9784	0.9092
Fat Content	Cubic	$Y = 1.37 - 0.0452A + 0.0955B - 0.0124AB - 0.0218A^2 - 0.0266B^2 - 0.0347A^2B - 0.0081AB^2 - 0.0349A^3 - 0.0047B^3$	$< 0.0037$	0.9917	0.9703
Protein Content	Cubic	$Y = 4.03 - 0.1405A - 0.0848B - 0.0341AB - 0.1760A^2 - 0.0410B^2 + 0.0220A^2B - 0.0389AB^2 - 0.3442A^3 - 0.0496B^3$	$< 0.0001$	0.9993	0.9972
Carbohydrates	Linear	$Y = 88.40 + 1.84A + 1.17B$	$< 0.0001$	0.8999	0.8562
Crude Fibers	Cubic	$Y = 1.75 + 0.0238A - 0.0018B - 0.0244AB + 0.0384A^2 + 0.0016B^2 - 0.0018A^2B - 0.0259AB^2 - 0.0780A^3 - 0.0483B^3$	$< 0.0390$	0.9280	0.7126
Phytochemical Content Total Phenol	Linear	$Y = 1410.49 - 262.34A - 113.06B$	$< 0.0001$	0.9746	0.9663
Flavonoid	Cubic	$Y = 85.45 - 4.95A - 0.5254B - 0.0793AB + 0.2632A^2 - 0.0565B^2 - 0.6959A^2B + 0.7383AB^2 - 0.3087A^3 - 0.9389B^3$	$< 0.0363$	0.9927	0.9738
Vitamin C	Quadratic	$Y = 5.55 - 0.2540A - 0.1778B - 0.0128AB - 0.0538A^2 - 0.0759B^2$	$< 0.0360$	0.9524	0.9047
Antioxidant Activity	Linear	$Y = 35.68 - 4.06A - 3.86B$	$< 0.0001$	0.8819	0.8392
Tannins	Cubic	$Y = 0.1009 - 0.0058A - 0.0006B - 0.0001AB + 0.0003A^2 - 0.0001B^2 - 0.0008A^2B + 0.0009AB^2 - 0.0004A^3 - 0.0011B^3$	$< 0.0363$	0.9927	0.9738



Table 5. Optimized response components, targets, limits, and interests in the formula optimization stage

Response Name	Goal	Lower Limit	Upper Limit	Lower Weight	Upper Weight	Importance
A: Temperature	In Range	50	65	1	1	3
B: Time	In Range	10	25	1	1	3
Yield	Maximize	28.17	38.40	1	1	3
Water	Maximize	4.45	9.77	1	1	5
Ash	Maximize	0.26	0.33	1	1	5
Fat	Maximize	1.20	1.46	1	1	5
Protein	Maximize	3.14	4.41	1	1	5
Carbohydrates	Maximize	84.16	90.80	1	1	5
Crude Fibers	Maximize	1.63	1.88	1	1	5
Total Phenol	Maximize	1043.18	1796.94	1	1	5
Flavonoid	Maximize	79.03	92.42	1	1	5
Vitamin C	Maximize	5.01	5.82	1	1	5
Antioxidant Activity	Maximize	27.18	41.96	1	1	5
Tannins	Maximize	0.09	0.11	1	1	5

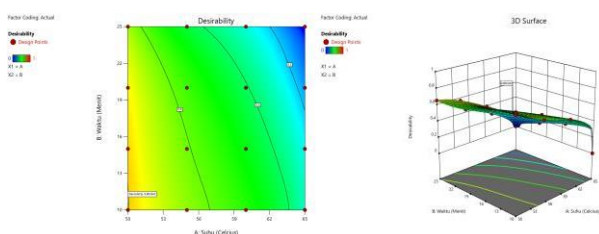


Fig. 13 Contour plot chart and response of optimum surface of corn bran

### 3.3. Model analysis for chemical and phytochemical testing in corn bran

Table 5 shows the optimized components, their targets, minimum and maximum, and the level of importance at the stage of the optimization formula. Optimization is then carried out by the method of optimal design. This method will optimize the corresponding variable data and the measurement data of the response is entered. The output of stage optimization is some recommended a new formula that is optimal according to the program Design Expert. Optimization is done by determining the limits (goal) criteria, the response is desired with a range that allows it to be achieved. The most optimal formula is the formula with the value of the maximum desirability. The value of desirability is the value function for optimization purposes, which shows the ability of the program to meet the desires based on the criteria outlined in the final product. The value of the desirability that is closer to a value of 1 indicates the ability of the program to produce the more desired product. In addition, the optimization is done after the quadratic mathematical model for each response, and the optimization is done to get the desired response (desirability).

Testing response and phytochemical is the component that is optimized with maximum target with 5 interest rate (+++++). Testing response and phytochemical is the response that determines the efficiency of the process. Time and temperature are optimized with a range of 3 interest rates (+++). The different levels of importance, essentially because of the maximized level of optimization.

Time and temperature will affect the quality of corn bran produced. This condition has a value of high desirability of 0.80 as shown in Figure 13. After the analysis of the model, performed the determination of the optimum temperature and time to produce the optimum conditions for testing and phytochemicals. The value of desirability that is produced is 0.80 (Montgomery, 2001). The optimum solution was obtained at a temperature of 50°C and 10 minutes. From the optimum solution, it is expected to produce yield at 38.34%, nutritional contents (water at 9.17%, ash at 0.33%, fat at 1.33%. protein at 4.40%, carbohydrate at 85.47%, and crude fiber at 1.88%) and phytochemical contents (total phenols at 1778.07 µg GAE/g. flavonoids at 92.11 µg GAE/g. vitamin C at 5.84 mg, antioxidant activity at 43.33%, and tannins at 0.11%).

## 4. CONCLUSION

The optimum conditions were obtained on the value of the desirability of 0.80 at a temperature of 50°C and 10 minutes. From the optimum solution, it is expected to produce yield at 38.34%, nutritional contents (water at 9.17%. ash at 0.33%. fat at 1.332%. protein at 4.40%. carbohydrate at 85.47%. and crude fibers at 1.88%), and phytochemical contents (total phenols at 1778.07 µg GAE/g. flavonoids at 92.11 µg GAE/g. vitamin C at 5.84 mg, antioxidant activity at 43.33%, and tannins at 0.11%).

## ACKNOWLEDGMENT

We are grateful to the Rector of Semarang University who has financially supported this research and facilitated by Research and Community Service Institutions Semarang University.

## REFERENCE

- [1] M. K. Sharif, M. S. Butt, F. M. Anjum, and S. H. Khan, "Rice Bran: A Novel Functional Ingredient," *Crit. Rev. Food Sci. Nutr.*, vol. 54, no. 6, pp. 807–816, 2014, DOI: 10.1080/10408398.2011.608586.
- [2] K. Gul, B. Yousuf, A. K. Singh, P. Singh, and A. A. Wani,

- "Rice bran: Nutritional values and its emerging potential for development of functional food - A review," *Bioact. Carbohydrates Diet. Fibre*, vol. 6, no. 1, pp. 24–30, 2015, DOI: 10.1016/j.bcdf.2015.06.002.
- [3] Isnawati and T. S. Mahanani, *Utilization of Agricultural Waste For the Production of Mycelium Beauveria bassiana*. Surabaya: Research Report. Research Institute Of Surabaya State University. Surabaya, 2003.
- [4] S. Widowati, "The utilization of the byproduct of Rice Milling in Supporting the System of Agro-industries in Rural areas," *AgroBio Bull.*, vol. 4, no. 1, pp. 33–38, 2001.
- [5] A. J. Henderson *et al.*, "Chemopreventive properties of dietary rice bran: Current status and prospects," *Adv. Nutr.*, vol. 3, no. 5, pp. 643–653, 2012, DOI: 10.3945/an.112.002303.
- [6] I. Pasha, F. Ahmad, Z. Siddique, and F. Iqbal, "Probing the effect of physical modifications on cereal bran chemistry and antioxidant potential," *J. Food Meas. Charact.*, vol. 14, no. 4, pp. 1909–1918, 2020, DOI: 10.1007/s11694-020-00438-9.
- [7] S. Maulana and Nurtahara, "Extraction of Phenol Compounds in the Liquid Smoke by Pyrolysis of Oil Palm Fronds," *J. Innov. Technol.*, vol. 1, no. 1, pp. 1–5, 2020, DOI: 10.31629/jit.v1i1.2127.
- [8] T. Charisma, "Studies of hypocholesterolemic rice analogs in vivo in Sprague-Dawley rats (SD)," Thesis at Bogor Agricultural Institute, 2015.
- [9] Y. Amir, S. Sirajuddin, and A. Levant, "Receptivity of Rice Bran Milk As A Functional Food," *Hasanuddin J. Public Heal.*, vol. 1, no. 1, pp. 16–25, 2020.
- [10] F. Kusnandar, S. Suryani, and S. Budijanto, "The characteristics of the Functional, Physical and Sensory Breakfast Corn Cereal Substituted With Rice Bran," *Appl. J. Food Technol.*, vol. 9, no. 3, pp. 108–117, 2020.
- [11] T. Khoerunisa, "Review: The Development Of Functional Food Products In The Indonesia-based on Superior Local Food Ingredients," *Indones. J. Agric. Food Res.*, vol. 2, no. 1, pp. 49–59, 2020.
- [12] M. Sholihah, U. Ahmad, and I. W. Budiastara, "Application of Ultrasonic Waves to Improve the Yield of Extraction and the Effectiveness of Antioxidants Mangosteen Skin," *J. Rancidity Agric.*, vol. 5, no. 2, pp. 161–168, 2017.
- [13] A. Handaratri and Y. Yuniati, "The Study of Extraction of Anthocyanins from the Mulberry Fruit by the Method of Sonication and Microwave," *Sci. J. Civ. Eng. Chem. Eng.*, vol. 4, no. 1, pp. 63–67, 2019.
- [14] Ibrahim, Yuniata, and Sriherfyna, "Effect of temperature and time of extraction on chemical and physical properties in making Zingiber officinale red ginger juice drinks var. Rubrum with a combination of the addition of honey as a sweetener," *J. Food Agro Ind.*, vol. 2, no. 2, pp. 530–541, 2015.
- [15] N. Yuliantari, I. Widarta, and I. Permana, "The influence of temperature and time of extraction on the content of flavonoids and the antioxidant activity of the soursop leaves (*Annona muricata* L.) using ultrasonic," *Sci. J. Food Technol.*, vol. 4, no. 1, pp. 35–42, 2017.
- [16] S. Abramson and A. K. Singh, "Treatment of the alcohol intoxications: Ethylene glycol, methanol, and isopropanol," *Curr. Opin. Nephrol. Hypertens.*, vol. 9, no. 6, pp. 695–701, 2000, DOI: 10.1097/00041552-200011000-00017.
- [17] F. Sebayang, "Pembuatan Etanol Dari Molase Secara Fermentasi Menggunakan Sel *Saccharomyces cerevisiae* Pada Kalsium Alginat," *J. Teknol. Proses*, vol. 5, no. 2, pp. 68–74, 2006.
- [18] K. K. Chew, S. Y. Ng, Y. Y. Thoo, M. Z. Khoo, W. M. Wan Aida, and C. W. Ho, "Effect of ethanol concentration, extraction time and extraction temperature on the recovery of phenolic compounds and antioxidant capacity of Centella Asiatica extracts," *Int. Food Res. J.*, vol. 18, no. 2, pp. 571–578, 2011.
- [19] D. E. Parasetia, Ritaningsih, and Hamsa, "The Making Natural Dyes From Jackfruit Wood," *J. Chem. Technol. Ind.*, vol. 1, no. 1, pp. 502–507, 2012.
- [20] Q. L. Hu, L. J. Zhang, Y. N. Li, Y. J. Ding, and F. L. Li, "Purification and anti-fatigue activity of flavonoids from corn silk," *Int. J. Phys. Sci.*, vol. 5, no. 4, pp. 321–326, 2010.
- [21] R. Hendryani, M. Lutfi, and L. C. Hawa, "The extraction of antioxidants dried leaves of red betel (*Piper scrotum*) by the method of pre-treatment ultrasonic-assisted extraction (the comparative study of the solvent type and the long time of extraction)," *J. Bioprocess Trop. Command.*, vol. 3, no. 2, pp. 33–38, 2015.
- [22] AOAC, "Association of Official of Analytical Chemistry," *Methods Anal. AOAC*, vol. 2, no. 2, pp. 148–148, 2007, [Online]. Available: <http://www.sciencedirect.com/science/article/pii/S0165993690870987>.
- [23] E. Candesa and L. Parker, *Handbook of Antioxidant*. New York: Marcell Dekker Inc, 2007.
- [24] J. Liu *et al.*, "Supercritical fluid extraction of flavonoids from maydis stigma and its nitrite-scavenging ability," *Adv. J. Food Sci. Technol.*, vol. 89, pp. 333–339, 2011, DOI: 10.19026/ajfst.6.3037.
- [25] I. Yinusa, H. A. Adeiza, and W. A. Balkees, "Phytochemical, Proximate, and Antimicrobial Analysis of the Fruit of *Sarcocephalus latifolius* (Smith Bruce)," *LAJANS Lapai J. Appl. Nat. Sci.*, vol. 1, no. 2, pp. 38–46, 2015.
- [26] C. S. Dzah *et al.*, "The effects of ultrasound-assisted extraction on yield, antioxidant, anticancer and antimicrobial activity of polyphenol extracts: A review," *Food Biosci.*, vol. 35, 2020, DOI: 10.1016/j.fbio.2020.100547.
- [27] L. Astráin-Redín, M. Alejandre, J. Raso, G. Cebrián, and I. Álvarez, "Direct Contact Ultrasound in Food Processing: Impact on Food Quality," *Front. Nutr.*, vol. 8, no. January, pp. 1–11, 2021, doi: 10.3389/fnut.2021.633070.
- [28] Haslina, N. Nazir, S. B. Wahjuningsih, and D. Larasati, "The influence of the type of solvent and extraction temperature of corn silk extracts," *Int. J. Adv. Sci. Eng. Inf. Technol.*, vol. 9, no. 3, pp. 911–915, 2019, DOI: 10.18517/ijaseit.9.3.9037.
- [29] S. Jamilatun, Y. Elisthatiana, S. N. Aini, I. Mufandi, and A. Budiman, "Effect of Temperature on Yield Product and Characteristics of Bio-oil From Pyrolysis of *Spirulina platensis* Residue," *Elkawmie*, vol. 6, no. 1, p. 96, 2020, DOI: 10.22373/ekw.v6i1.6323.
- [30] A. M. Pannase, R. K. Singh, B. Ruj, and P. Gupta, "Decomposition of polyamide via slow pyrolysis: Effect of heating rate and operating temperature on product yield and composition," *J. Anal. Appl. Pyrolysis*, vol. 151, 2020, DOI: 10.1016/j.jaap.2020.104886.
- [31] H. Haslina, N. Nazir, and A. Sampurno, "Different Drying Duration of Corncobs Powders and Its Effects on Physical, Nutritional and Phytochemical," *Int. J. Adv. Sci. Eng. Inf. Technol.*, vol. 11, no. 3, pp. 1232–1238, 2021, DOI: 10.18517/ijaseit.11.3.15321.
- [32] S. E. Uwadiae, H. Aifesome, and B. V. Ayodele, "Effect of extraction temperature, time and volume of diluent on oil yield from ginger (*Zingiber officinale*) in a batch-mode process," *J. Appl. Sci. Environ. Manag.*, vol. 23, no. 4, p. 611, 2019, DOI: 10.4314/jam.v23i4.5.
- [33] S. Fitriani, "Pengaruh Suhu dan Lama Pengeringan Terhadap Beberapa Mutu Manisan Belimbing Wuluh (*Averrhoa bilimbi* L.) Kering," *Sagu*, vol. 7, no. 1, pp. 32–37, 2008.
- [34] H. H. S. Abdel-Naeem, K. I. Sallam, and H. M. B. A. Zaki, "Effect of different cooking methods of rabbit meat on topographical changes, physicochemical characteristics, fatty acids profile, microbial quality and sensory attributes," *Meat Sci.*, vol. 181, no. November 2021, 2021, doi: 10.1016/j.meatsci.2021.108612.
- [35] T. Anukiruthika, J. A. Moses, and C. Anandharamakrishnan, "Electrohydrodynamic drying of foods: Principle, applications, and prospects," *J. Food Eng.*, vol. 295, 2021, DOI: 10.1016/j.jfoodeng.2020.110449.
- [36] B. Z. Zaki, S. Appalasamy, M. M. Nor, and A. Eh Rak, "Effect of Temperature on Moisture, Ash and Crude Fat Content in Etak (*Corbicula fluminea*) Tissue via Modified Oven Smoking Method," in *IOP Conference Series: Earth and Environmental Science*, 2020, vol. 549, no. 1, DOI: 10.1088/1755-1315/549/1/012056.
- [37] M. Darmawan, Syamdidi, and E. Hastarini, "Pengolahan Bakto Agar dari Rumpun Laut Merah (*Rhodomyenia ciliata*) dengan Pra Perlakuan Alkali," *J. Pascapanen dan Bioteknologi Kelaut. dan Perikan.*, vol. 1, no. 1, p. 9, 2006, doi: 10.15578/jpbkp.v1i1.83.
- [38] S. Kumala, R. Sumarni, R. Rachmani, and A. Ruswita, "Alga Merah (*Gracilaria verrucosa*) sebagai Bahan Bakto Agar," *J. Farm. Indones.*, vol. 6 No. 3, no. 3, pp. 166–171, 2013.
- [39] T. Czaja, A. Sobota, and R. Szostak, "Quantification of ash and moisture in wheat flour by Raman spectroscopy," *Foods*, vol. 9, no. 3, p. 280, 2020, DOI: 10.3390/foods9030280.
- [40] G. G. Hewavitharana, D. N. Perera, S. B. Navaratne, and I. Wickramasinghe, "Extraction methods of fat from food samples

- and preparation of fatty acid methyl esters for gas chromatography: A review," *Arab. J. Chem.*, vol. 13, no. 8, pp. 6865–6875, 2020, DOI: 10.1016/j.arabjc.2020.06.039.
- [41] S. Ismiyanto, A. Halim, and P. J. Wibawa, "Identifikasi Komposisi Asam Lemak dari Minyak Benih," *J. Kim. Sains Apl.*, vol. 9, no. 1, pp. 1–5, 2006.
- [42] S. P. S. He, Nurjanah, and A. M. Jacob, "Chemical Composition And Antioxidant Activity Of Root, Stem Bark And Lindur Leaves," *Process. Results Indones. Fish.*, vol. 18, no. 2, pp. 205–219, 2015, [Online]. Available: <https://core.airconditioning.uk/download/pdf/291863573>
- [43] Q. Jiang, J. Han, P. Gao, L. Yu, Y. Xu, and W. Xia, "Effect of heating temperature and duration on the texture and protein composition of bighead carp (*Aristichthys Nobilis*) muscle," *Int. J. Food Prop.*, vol. 21, no. 1, pp. 2110–2120, 2018, DOI: 10.1080/10942912.2018.1489835.
- [44] M. Sajib, E. Albers, M. Langeland, and I. Undeland, "Understanding the effect of temperature and time on protein degree of hydrolysis and lipid oxidation during ensilaging of herring (*Clupea harengus*) filleting co-products," *Sci. Rep.*, vol. 10, no. 1, 2020, DOI: 10.1038/s41598-020-66152-0.
- [45] F. Nurani, T. Dhalika, and A. Budiman, "Mekanisme Produksi Protein Asal Daun Singkong (*Manihot Utilisima*) Sebagai Bahan Pakan Dengan Menggunakan Metode Pelarutan Pada Suhu Yang Berbeda," *J. Univ. Padjadjaran*, vol. 5, no. 1, pp. 1–10, 2016.
- [46] Yulianti, B. Susilo, and R. Yulianingsih, "Pengaruh lama ekstraksi dan konsentrasi pelarut etanol terhadap difat fisika-kimia ekstrak daun stevia (*Stevia rebaudiana bertonii* M.) dengan metode microwave assisted extraction (MAE)," *J. Bioproses Komod. Trop.*, vol. 2, no. 1, pp. 35–41, 2014.
- [47] O. W. Nilasari, W. H. Susanto, and J. M. Maligan, "Pengaruh Suhu dan Lama Pemasakan Terhadap Karakteristik Lempok Labu Kuning (Waluh)," *J. Pangan dan Agroindustri*, vol. 5, no. 3, pp. 15–26, 2017.
- [48] B. Jinturkar, "The effect of temperature and carbohydrates sources on the growth of rhizobium," *Int. J. Dev. Res.*, vol. 7, no. 6, pp. 13302–13303, 2017.
- [49] M. A. Scariot, L. Karlinski, R. G. Dionello, A. L. Radünz, and L. L. Radünz, "Effect of drying air temperature and storage on the industrial and chemical quality of rice grains," *J. Stored Prod. Res.*, vol. 89, 2020, DOI: 10.1016/j.jspr.2020.101717.
- [50] H. Huang *et al.*, "Modification of tea residue dietary fiber by high-temperature cooking assisted enzymatic method: Structural, physicochemical and functional properties," *Lwt*, vol. 145, 2021, DOI: 10.1016/j.lwt.2021.111314.
- [51] M. Hernanz *et al.*, "Extraction of phenolic compounds from the cocoa shell: Modeling using response surface methodology and artificial neural networks," *Sep. Purif. Technol.*, vol. 270, 2021, DOI: 10.1016/j.seppur.2021.118779.
- [52] M. E. Setyantoro, H. Haslina, and S. B. Wahjuningsih, "Pengaruh Waktu Ekstraksi Dengan Metode Ultrasonik Terhadap Kandungan Vitamin C, Protein, dan Fitokimia Ekstrak Rambut Jagung (*Zea mays* L.)," *J. Teknol. Pangan dan Has. Pertan.*, vol. 14, no. 2, pp. 53–67, 2019, doi: 10.26623/jtphp.v14i2.2445.
- [53] X. Chen *et al.*, "Effect of blanching and drying temperatures on starch-related physicochemical properties, bioactive components and antioxidant activities of yam flours," *LWT - Food Sci. Technol.*, vol. 82, pp. 303–310, 2017, DOI: 10.1016/j.lwt.2017.04.058.
- [54] S. Lenny, *Senyawa flavonoid, fenilpropanoida, dan alkaloida*. Medan: Fakultas Matematika dan Ilmu Pengetahuan Alam (MIPA) Universitas Sumatera Utara (USU), 2006.
- [55] L. Qiao *et al.*, "Sonochemical effects on 14 flavonoids common in citrus: Relation to stability," *PLoS One*, vol. 9, no. 2, 2014, DOI: 10.1371/journal.pone.0087766.
- [56] Seema and U. Yadava, "Study of the heating effect on the vitamin C of citrus fruits," *Int. J. Food Allied Sci.*, vol. 3, no. 1, pp. 36–42, 2017.
- [57] I. E. Shara and S. Ben Mussa, "Determination of Vitamin C (Ascorbic Acid) Contents in Vegetable Samples by UV-Spectrophotometry and Redox Titration Methods and Estimation the Effect of Time, Cooking and Frozen on Ascorbic Acid Contents," *International Journal of Progressive Sciences and Technologies (IJPSAT)*, vol. 15, no. 2, pp. 281–293, 2019, [Online]. Available: <http://ijpsat.ijst-journals.org>.
- [58] G. Ssepuuya, D. Nakimbugwe, A. De Winne, R. Smets, J. Claes, and M. Van Der Borgh, "Effect of heat processing on the nutrient composition, color, and volatile odor compounds of the long-horned grasshopper *Ruspolia different serville*," *Food Res. Int.*, vol. 129, 2020, DOI: 10.1016/j.foodres.2019.108831.
- [59] G. Hao *et al.*, "Effect of temperature on chemical properties and antioxidant activities of abalone viscera subcritical water extract," *J. Supercrit. Fluids*, vol. 147, no. 2019, pp. 17–23, 2019, DOI 10.1016/j.supflu.2019.02.007.
- [60] F. Kosakih, "The influence of Solvent Type, Temperature and Extraction Time On the Antioxidant Activity of the Extract of the Soursop Leaves (*Annona muricata* L.) and Its Application in Hard Candy Products," Semarang, 2017.
- [61] Badriyah, D. J. Ahmadi, and L. K. Nuswantara, "In Vitro Rumen Degradability of Phenolic Compound and Antioxidant Activity of *Moringa oleifera* Leaf," *Indones. Livest. J.*, vol. 19, no. 3, pp. 116–121, 2017.
- [62] S. Ogawa, "Studies on Antioxidant Activity in Japanese Edible Seaweeds," Tokyo University of Fisheries, Tokyo, 2003.
- [63] A. S. C. Teles, D. W. H. Chávez, F. Dos Santos Gomes, L. M. C. Cabral, and R. V. Tonon, "Effect of temperature on the degradation of bioactive compounds of Pinot Noir grape pomace during drying," *Brazilian J. Food Technol.*, vol. 21, p. 6723 5917, 2018, DOI: 10.1590/1981-6723.5917.